Review The Notch3 Pathway in Organ Fibrosis

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Received: 19 September 2024; Accepted: 21 October 2024; Available online: 23 October 2024

ABSTRACT: Fibrosis occurs in many organs, including the lung, heart, skin, liver or kidney, and is characterized by progressive tissue scarring in response to repetitive or chronic non-resolving injury, ultimately leading to organ failure and death. It is, in fact, a major cause of morbidity and mortality worldwide, being estimated to account for 45% of deaths in the world. Despite this fact, little progress has been made therapeutically, and fibrosis remains a major clinical and therapeutic challenge. Although significant advances in our understanding of cellular and molecular mechanisms driving tissue fibrosis have been made, the lack of an efficient treatment reflects the limited insight into the pathophysiological mechanisms underlying the initiation and progression of the fibrotic process. Thus, there is an urgent need for better understanding of tissue fibrosis and repair mechanisms that later lead to the development of new therapeutic approaches to fight fibrosis. The Notch pathway is a highly conserved signaling pathway that has been linked to tissue fibrosis in many organs and promises to open new therapeutic opportunities. This manuscript reviews the relevance of Notch signaling in the development and progression of tissue fibrosis in several organs with a special focus on the Notch3 pathway due to the unique features of this receptor.

Keywords: Tissue fibrosis; Notch pathway; Intercellular communication; Fibroblast activation; Collagen deposition

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1. Introduction

Fibrosis is a biological process that leads to extracellular matrix (ECM) deposition with the purpose of giving the tissue the biological scaffold, whereby it can repair properly after injury. However, factors such as repetitive tissue injury and/or aberrant healing response may lead to pathological fibrosis, where ECM deposition does not terminate and accumulates, leading to tissue scarring and organ failure [1–3]. Fibrotic diseases are estimated to be present in around 45% of all deaths worldwide [4]. Indeed, fibrosis can affect many organs, including the liver, kidney, skin, heart, and lung [5]. Unfortunately, despite the huge morbidity, mortality and economic impact, the few antifibrotic therapies available in the clinical setting usually lead to adverse effects. In many cases, they just slow down disease progression [3], leaving a tremendous need for the development of new therapies that cure organ fibrosis. Fibroblasts play a major role in the fibrotic process. In response to multiple signals, fibroblasts activate and differentiate into myofibroblasts, acquiring a contractile phenotype similar to that of smooth muscle cells, showing apoptosis resistance and producing exaggerated amounts of collagen and other ECM proteins [3,6,7]. Many soluble factors and signaling cascades have been involved in these processes [7]. Unfortunately, although some of the cellular and molecular mechanisms promoting pathological fibrosis are shared among organs, there is also an evident variability, posing a huge challenge towards finding therapeutic targets to prevent, halt the progression, or revert tissue scarring.

Numerous signaling cascades have been involved in mediating fibrogenesis. One such mechanism is the Notch pathway, which, contrary to other pathways, has not been deeply explored in tissue fibrosis. Thus, this manuscript reviews what is known about the role of Notch signaling in the development and progression of tissue fibrosis in different organs, with a particular focus on Notch3 signaling.

2. Notch Pathway in Tissue Fibrosis

The Notch pathway is a highly conserved signaling pathway involved in many different biological processes, such as developmental patterning, cell fate acquisition, cell apoptosis, proliferation, survival and tissue regeneration [8,9]. The *Notch* gene was first identified in *Drosophila melanogaster* more than 100 years ago, and its name comes from its role in the specific phenotype displayed by the notched wings in these flies [10]. However, while *Drosophila* has a single Notch gene, gene duplication and diversification during evolution have given rise to four Notch paralogs in mammals (Notch1-4). These genes produce four different Notch proteins (Notch1-4), displaying overlapping or specific functions, depending on the biological context. Notch is a transmembrane protein, and so are the five canonical Notch ligands: Jagged (Jag) 1, Jag2, Delta-like ligand 1 (Dll1), Dll3, and Dll4. The molecular structure of Notch receptors and ligands has been well described (Figure 1A), showing both similarities and differences among Notch receptors and ligands. The canonical Notch pathway is a signaling cascade that mediates direct cell-cell physical interactions. The receiver cell shows the receptor in the membrane, and the neighbour cell exhibits the ligand. Upon direct binding of the ligands to Notch receptors, the released Notch intracellular domain (NICD) derived after cleavage translocates into the nucleus, where it binds to CBF-1/supressor of hairless/Lag1 (CSL, also called recombination signal binding protein-J, RBPJk) to induce transcription of the Hes and Hey family target genes to regulate distinct biological processes aforementioned. Importantly, the Notch pathway is a cell and context-dependent signaling cascade. Thus, its role varies in specific circumstances depending on the cell type and the biological context. In addition, apart from being a complex pathway itself, the Notch pathway has also been described to interact with various pathways such as TGFβ [11], YAP/TAZ [12], the Sonic Hedgehog pathway [13], and Wnt [13], which broadens the amount of different possible outcomes related to Notch pathway activation.

Figure 1. The Notch receptors. (A), The four Notch receptors with their main structural features depicted. (B), The Notch3 receptor and its unique molecular features are represented. N1, N2, N3, N4: Notch1, Notch2, Notch3, Notch4; NECD: Notch extracellular domain; TMD: Transmembrane domain; NICD: Notch intracellular domain.

3. Implication of Notch3 in the Development of Organ Fibrosis

As mentioned above, the four Notch receptors share a similar basic structure but differ in several aspects. The Notch receptors have a similar extracellular domain, which contains three lin-12/Notch /LNR) motifs, 29–36 EGF-like repeats, and a heterodimerization domain. While Notch1 and Notch 2 have 36 EGF-like repeats, Notch 3 has 34 and Notch4 29, resulting in Notch3 having a shorter ECD than Notch1 and Notch2 but longer than Notch4 ECD [14] (Figure 1). Moreover, Notch3 and Notch4 lack the transactivation domain (TAD) found in the N1ICD and N2ICD (Figure 1A). In contrast, Notch3 has a unique PPXY motif, known to act as a WW domain recognition site for the endocytic regulator WWP2 (Figure 1B). Furthermore, Notch4 lacks the Notch cytokine response (NCR) region present in the other Notch receptors [15] (Figure 1). All of these structural differences in the Notch receptors may account for the functional differences observed among them. Interestingly, structural data has shown that Notch3 seems to be the most easily cleaved of all Notch receptors [16]. In fact, data shows that it can be cleaved and activated even in the absence of a ligand [17]. Furthermore, Notch3 has specific mutational hotspots that are associated with the development of Cerebral Autosomal Dominant Arteriopathy with Subcortical Infarcts and Leukoencephalopathy (CADASIL), a hereditary neurological disorder characterized by vascular abnormalities in the brain [18]. This is the major implication of Notch3

dysfunction in human disease. In addition, it has been shown to be involved in several cancers including breast, lung, pancreatic, and colorectal cancer [19], and in fibrotic disease in distinct tissues, which is described in detail next.

3.1. Notch3 in Liver Fibrosis

Continuous liver injury may lead to extensive liver fibrosis, also known as cirrhosis, which is a major cause of death worldwide [20]. The Notch pathway plays an important role in liver development and regeneration [21]; therefore, a dysregulation in this signaling may give rise to the development of pathology.

Hepatic stellate cells (HSCs) are a type of resident mesenchymal cell that has features of both resident fibroblasts and pericytes and produces an extracellular matrix (ECM) [22]. HSCs activation into proliferative, fibrogenic myofibroblasts is considered the central driver of hepatic fibrosis in experimental and human liver injury [23]. In fact, single-cell RNA sequencing (scRNAseq) studies have shown that HSCs are the main source of myofibroblasts during cirrhosis [24]. Macrophages are found in close proximity to activated HSCs and have been demonstrated to play a key role in fibrosis initiation and progression [25,26]. Both Kupffer cells (resident macrophages) and recruited macrophages produce pro-fibrotic mediators such as transforming growth factor β (TGFβ) and platelet-derived growth factor (PDGF), cytokines and chemokines that further lead to worsened fibrosis [26,27]. Interestingly, Notch signaling has been previously shown to be crucial for HSC activation and macrophage polarization, determining the fate of liver fibrogenesis [28].

The specific implication of Notch3 signaling in liver fibrosis has been previously described by Chen et al. [29]. They found that the Notch3 pathway could regulate the activation of HSCs. They observed by immunohistochemistry that all fibrotic liver tissues from patients with chronic active hepatitis were positive for Notch3, whereas Notch3 was not detected in normal liver tissues. Futhermore, they showed that overexpression of Notch3 led to increased expression of α-SMA and collagen I in HSC-T6 cells, while downregulation of Notch3 yielded the opposite effects. Consistently, the expression of Notch3 has been found to be upregulated in diseased human livers and a rat model of liver fibrosis [30,31]. In contrast, in a study using a mouse model of liver fibrosis, the authors could not detect changes in Notch3, but rather an increase in the expression and activation of Notch1 and Notch2 [32]. However, multi-lineage modelling of ligand and receptor interactions has revealed Notch3 signaling as an important regulator of mesenchymal cell function within the human liver fibrotic niche [33]. This study, describes the interaction between Notch ligands derived from scar-associated endothelial cells and Notch3+ scar-associated mesenchymal cells. Indeed, coculture of primary human HSCs and endothelial cells from cirrhotic liver promoted fibrillar collagen production by HSCs, which was inhibited using a Notch-signaling inhibitor. Accordingly, the knockdown of Notch3 expression in primary human HSCs resulted in reduced fibrillar collagen expression. In addition, Cong and Liu et al. reported that Notch3 expression at the mRNA and protein levels is significantly higher in patients with moderate and severe liver fibrosis compared with patients without liver fibrosis. In contrast, no significant difference existed between mild patients and patients without liver fibrosis [34]. This study, also showed that the fibrogenic factors such as α-SMA, collagen I, TGF-β1 and Smad3 were significantly increased upon Notch3 upregulation in a human HSC line. Accordingly, the opposite effect was observed when Notch3 was downregulated. In addition, the Notch target gene Hes1 was increased in liver biopsies from patients with nonalcoholic steatohepatitis (NASH)/fibrosis [32], suggesting a potential implication of the Notch3/Hes1 axis in hepatic fibrosis.

Based on the ample evidence of the regulatory role of the Notch pathway in liver fibrosis, its modulation emerges as a potential novel therapy for cirrhotic patients to improve symptoms or even revert the disease.

3.2. Notch3 in Kidney Fibrosis

Chronic kidney disease (CKD) affects more than 10% of the world's population, and its progression may lead to renal fibrosis [35]. Notch3 is considered one of the major receptors involved in renal fibrosis [36] despite the fact that few studies have established its precise role in this context. Djudjaj et al. revealed in 2012 the likely critical implication of Notch3 in kidney fibrosis since genetic deletion of Notch3 using global Notch3-knockout mice protected them from tubulointerstitial fibrosis induced by unilateral ureteral obstruction (UUO), a murine model of kidney fibrosis, exhibiting significantly lower collagen deposition and reduced number of α -SMA-positive cells [37]. Furthermore, the Notch target genes HeyL, Hes5, and Hey2 were shown to be potentially downstream of Notch3 signaling in this context. In a study by Xiao et al. [38], the authors showed that the expression of Notch 1, 3, and 4, Notch intracellular domain (NICD), and its target genes Hes1 and HeyL were upregulated in mice with UUO. Importantly, pharmacological

inhibition using the pan-Notch inhibitor DBZ resulted in a reduction of pathological fibroblasts and other fibrotic parameters, indicating the relevant implication of Notch signaling in kidney fibrogenesis.

In addition to myofibroblasts, which are considered the major cellular player in the development and progression of kidney fibrosis, other cell types also contribute; this is the case of tubule epithelial cells, in which sustained activation of the Notch pathway has a key role in fibrogenesis [39]. Interstitial inflammation and fibrosis are one of the major pathological changes of polycystic kidney disease (PKD), a genetic disorder affecting millions of people worldwide [40]. A recent study observed that Notch3 was highly upregulated in patients with PKD [41]. These authors showed that chronic epithelial overexpression of N3ICD in mice resulted in severe tubulointerstitial inflammation and fibrosis that progressed with time. Interestingly, Notch3 deficiency may also predispose to kidney fibrosis, as indicated by anecdotal reports from CADASIL patients [42] and research involving angiotensin II-infused mice [43]. By contrast, one study shows that the global deletion of Notch3 did not protect mice from fibrosis, claiming that Notch3 minimally contributes to CKD [44]. A recent study showed that Notch3 expression on CD45+ leucocytes regulates immune cell infiltration following UUO [45]. Interestingly, using chimeric animal models, fibrosis still ensued despite the lack of prominent leukocyte infiltrates. However, using global Notch3 KO mice showed a reduction in renal fibrosis, demonstrating that Notch3 activation in kidney cells is crucial for the development of kidney fibrosis regardless of the presence of leukocyte infiltrates.

All these observations support a model in which the Notch3 pathway is reactivated in response to kidney injury and contributes to the fibrotic response, suggesting that decreasing Notch3 activity in CKD patients could potentially be used to ameliorate disease development.

3.3. Notch3 in Skin Fibrosis

Skin fibrosis has a wide variety of pathological manifestations such as hypertrophic scarring, keloid formation, chronic cutaneous Graft-versus-Host-Disease (GvDH), nephrogenic fibrosing dermopathy or systemic sclerosis (SSc) [46]. Nevertheless, the exact mechanisms leading to fibrotic skin conditions remain to be elucidated.

In vivo experiments using a skin incision model have shown that RBPJk KO mice exhibit a reduced expression of several collagens and fibrotic markers in the healed skin [47]. Consistently, pharmacological Notch inhibition by treatment with the gamma secretase inhibitor DAPT reduced bleomycin-induced fibrosis in a dose-dependent manner with significant decreases in dermal thickening, numbers of myofibroblasts and hydroxyproline content [48]. However, to our knowledge, there are no existing studies investigating specifically the direct effect of Notch3 signaling on the development or progression of skin fibrosis. A study focused on hair outgrowth and skin fibrosis revealed that the expression of Notch3 was elevated in αSMA+ dermal sheath cells when βcatenin was overexpressed, alongside the expression of fibrotic markers such as *Ctgf*, *Collal*, and *Colla2* [49].

Specific NOTCH3 polymorphisms correlate with the susceptibility to diffuse cutaneous systemic sclerosis (DcSSc) [50]. Treatment of human neutrophils, dermal microvascular endothelial cells, and primary skin fibroblasts with dSsc neutrophil-derived exosomes induced the expression of Notch pathway genes and fibrotic-related genes [51]. Recessive dystrophic epidermolysis bullosa (RDEB) is a rare skin fragility disorder caused by mutations in COL7A1, and progressive fibrosis is one of its major hallmarks. RDEB-derived fibroblasts treated with DAPT showed a reduction of fibrotic traits. However, in this case, it is probably due to the blockade of the Notch1 receptor rather than Notch3 due to the observed upregulation of the first one in these cells [52].

Interestingly, Notch3 has been demonstrated to induce terminal differentiation of keratynocites [53], the major cell type of the epidermis, and this could be used in treatment strategies focused on tissue repair. Overall, the general lack of studies of Notch signaling in skin fibrosis shows that there is plenty of scope for the study of Notch3 signaling in this pathology.

3.4. Notch3 in Heart Fibrosis

Cardiovascular disease is the leading cause of death worldwide, and cardiac fibrosis is a common pathophysiological manifestation of most cardiovascular diseases [54,55]. To our knowledge, the first observation of the potential role of Notch3 in cardiac fibrosis was reported in 2012 [56]. Here, they observed that apelin-13, a specific ligand for the angiotensin-like 1 receptor, upregulated the expression of several genes, including Notch3, and attenuated cardiac fibrosis in infarcted mice. This was accompanied by increased bone marrow cells (BMCs) recruitment. In addition, the treatment of cultured BMCs with apelin in vitro increased *Notch3* expression [57]. Further, treatment with bone marrow cells overexpressing apelin in vivo, significantly increased angiogenesis and attenuated cardiac fibrosis formation in post-MI mice, although whether this effect is mediated by Notch3 remains unknown [57].

A recent study reported that Notch3^{$-/-$} mice exhibit left ventricular hypertrophy combined with mild fibrosis [58]. Furthermore, it was shown that *Notch3* was expressed in vascular smooth muscle cells and pericytes and not in cardiomyocites, thereby suggesting that the observed phenotype could be due to a perturbation of Notch3 signaling in these cells. In line with this, Chen et al. indicated that a reduction in Notch3 may disrupt endothelial cell-pericyte communication, causing pericyte detachment that promotes its differentiation into myofibroblasts during ischemiareperfusion or myocardial infarction [59]. However, the exact mechanism by which Notch3 may be implicated in the development of cardiac fibrosis was not assessed [60].

Interestingly, a previous study showed that Notch3 overactivation in cardiac fibroblasts in vitro promotes apoptosis and inhibits proliferation, fibroblast to myofibroblast differentiation, and ECM production, while Notch3 downregulation shows the opposite effects [60]. Accordingly, overactivating Notch3 signaling in vivo further prevented miocardial infarction-induced cardiac fibrosis in rats [60]. Consistently, a similar study performed in mice demonstrated that Notch3 signaling inhibits TGF-β1/Smad3 signaling in cardiac fibroblasts, impairing fibroblast-to-myofibroblast transition, evidenced by a decrease in αSMA and Type I collagen expression [61]. Altogether, in contrast to the pathogenic role of Notch3 signaling upregulation observed in fibrosis in other organs, evidence support Notch3 downregulation as a driver of fibrosis in the heart and its activation as a therapeutic option to treat cardiac fibrosis.

3.5. Notch3 in Pancreatic Fibrosis

Pancreatic fibrosis is a characteristic feature of chronic pancreatitis and pancreatic cancer [62]. Even though, up to date, there is no direct association between Notch3 signaling and pancreatic fibrosis nor any published work focused on this topic, a couple of studies have observed a relation between Notch3 and chronic pancreatitis [63,64]. First, transcripts of Notch3 in microdissected ectatic ducts of chronic pancreatitis were found to be elevated [63]. In a recent study, the authors identified Notch3 as a risk gene and promising biomarker for chronic pancreatitis, which needs further confirmation in a clinical study [64]. Pancreatic cancer is one of the most lethal cancer types, and its incidence is increasing [65]. Pancreatic ductal adenocarcinoma (PDAC) is the most common type of pancreatic cancer and is characterized by the presence of abundant desmoplastic stroma. Desmoplasia is the formation of dense fibrotic tissue within and around tumor tissue, which is generated by activated fibroblasts, myofibroblasts, pancreatic stellate cells and tumor cells. It comprises 80% to 90% of the tumor volume [66]. Activated pancreatic stellate cells (PaSCs) are the key cellular source of cancer-associated fibroblasts in the pancreatic stroma of PDAC patients. Interestingly, Song et al. showed that Notch3 is overexpressed in both human stromal cells from PDAC patients and activated mouse PaSCs [67]. Remarkably, upon Notch3 knockdown in vitro, a decrease in both RNA and protein levels of the fibrotic markers αSMA, collagen I, and fibronectin, and also of the Notch target gene Hes1, was observed. Furthermore, cell proliferation and migration were reduced when Notch3 was downregulated [68]. All these findings suggest that Notch3 regulates cell migration, proliferation and expression of fibrotic markers in the stromal component of pancreatic cancer. Importantly, Notch3 expression inversely correlates with response to therapy and overall survival in patients with pancreatic cancer treated with gemcitabine [68], highlighting the relevant implication of this pathway in the disease. Thus, targeting Notch3 could impact pancreatic tumorigenesis by limiting fibrosis and emerge as a novel therapy for PDAC patients.

3.6. Notch3 in Lung Fibrosis

Idiopathic pulmonary fibrosis (IPF) is the most prevalent lung fibrotic disease within the interstitial lung diseases, a group of more than 200 parenchymal pulmonary disorders mainly characterized by alveolar damage and interstitial inflammation and/or fibrosis. IPF is a chronic and progressive pulmonary disease, and the fact that there is currently no cure, a lack of treatments impeding its progression and that patients are not always eligible to receive a lung transplant often leads to death in 3–5 years post-diagnosis. One of the histopathological features of IPF is the presence of fibroblasts foci, which are active sites of remodelling in the fibrotic lung where pathological fibroblasts produce and secrete excessive amounts of ECM proteins and are lined by aberrant epithelial cells not normally found in healthy lungs and with an important role in tissue fibrosis.

A study from 2010 showed that myofibroblast differentiation was impaired in vivo in Notch2−/−; Notch3−/− double mutant embryos but not in single mutants in the developing lung mesenchyme, suggesting the redundant function of these two receptors inducing myofibroblast differentiation [69]. In the adult lung, Notch1 plays an essential role in fibroblast-to-myofibroblast differentiation in the pathogenesis of pulmonary fibrosis, as reported by Hu et al. [70].

Interestingly, a previous publication identifies a unique Pdgfrβ+ population defined by Notch3 as a discriminating marker to emerge in the fibrotic mouse lung [71]. Furthermore, an upregulation of Notch3 in fibroblasts was observed by scRNA sequencing comparing healthy and fibrotic human lungs [72]. Our previous work demonstrates the specific role of Notch3 in fibroblast-to-myofibroblast differentiation [73]. First, we observed that around 60% of the collagenexpressing cells showed Notch3 activity in the mouse lung at homeostasis. After bleomycin-induced lung injury, almost half of the emerging αSMA+ myofibroblasts showed Notch3 activity, suggesting a role of Notch3 in fibrogenesis. To test this hypothesis, we used a Notch3 KO mouse line and observed that Notch3 deficiency attenuated bleomycininduced lung fibrosis and impeded lung function decline. Furthermore, we demonstrated that Notch3 signaling regulates fibroblast survival and myofibroblast differentiation. Consistently, a previous study showed that ROS-dependent activation of p38, JNK1/2, and Notch3 activation in IMR-90 cells (primary human lung fibroblasts) promoted basal and TGFβ1-induced differentiation and expression of ECM proteins in vitro [74]. Moreover, treatment with the Notch inhibitor DAPT or Notch3-specific siRNA suppressed the expression of α SMA. Using immunohistochemistry, the authors found Notch3 in the cytoplasm, membrane, and nuclei of myofibroblasts and in epithelial cells of IPF lungs, while they did not detect it in healthy lungs. In contrast, we detected Notch3 activity in α SMA+ myofibroblasts of the healthy lung and observed an expansion of the N3ICD+ myofibroblasts in the IPF lung [73]. All in all, targeting Notch3 signaling could be a potential strategy to tackle IPF progression.

4. Current Clinical Trials Using Notch Inhibitors

Filtering for "Clinical Trial" or "Randomized Controlled Trial" and searching for articles containing both terms "Notch3" and "fibrosis," yields zero results using PubMed as of September 2024. Similarly, when entering "fibrosis" as a condition/disease and "NOTCH inhibitor" as intervention/treatment in "clinicaltrials.gov", no results are obtained. However, when searching just for "NOTCH inhibitor", regardless of the condition/disease, a total of 50 studies are listed, suggesting that there is a significant unexplored venue for the establishment of new or repurposed drugs targeting Notch signaling for the treatment of fibrotic diseases.

The use of Notch inhibitors to treat cancer is extensive. Currently, two recruiting clinical trials use pan-NOTCH inhibitors in cancer (NCT05774899, NCT04973683). There are also three additional active studies with the same scope that are not recruiting (NCT02069730, NCT03785964, NCT04871282), which are also focused on the use of pan-NOTCH inhibitors in cancer and, interestingly, the two last are the only clinical trials in phase III. The rest of the clinical trials are also solely focused on treating different cancer diseases and are in phase I or II, with no clinical trials in phase IV. Of note, 22 of the 50 total trials have been terminated, meaning the study was stopped earlier and will not start again. While some of the studies did not specify the reason for termination (NCT00100152, NCT01088763, NCT01071564, NCT01193881, NCT01192763, NCT01238133, NCT01208441), others did: "Business reason" (NCT03422679), "Non-safety reason, business objectives have changed" (NCT01986218), "Administratively Complete" (NCT01151449, NCT01120275, NCT01269411), "Drug was no longer available" (NCT01200810), "Sponsor's decision" (NCT04461600), "Study drug production halted" (NCT01193868), "stopped prematurely by Company due to decision to terminate all CTEP supplied drug for further development of RO4929097" (NCT01122901), "Company decided to stop the development of drug" (NCT01189240), "The trial was closed because the sponsor became insolvent" (NCT03740100), and "Slow accrual coupled with discontinuation of study drug" (NCT01217411). Furthermore, the use of general Notch inhibitors often leads to adverse effects, which poses a huge problem in the establishment of these drugs as approved disease treatments.

There are some drugs related to Notch ligands that are currently being tested. For instance, two T-cell engagers targeting DLL3, a canonical Notch ligand, are being tested by Boehringer Ingelheim (BI764532) and Merck (HPN328) to treat lung cancer. Unfortunately, to our knowledge, no drugs that specifically inhibit Notch3 signaling are being tested.

The use of Notch3 neutraling antibodies has been proposed in several studies to treat different diseases. In vitro, α-Notch3 (R&D Systems, Minneapolis, MN, USA) has also been used to study the role of Notch3 in collagen II-specific T helper type 1 (Th1) and Th17-type expansion, proposing that selective inhibition of Notch signaling transduced by Notch3 could be used for the treatment of rheumatoid arthritis. More than a decade ago, the development of anti-Notch3 monoclonal antibodies to either block (A4 and A8) or activate (A45 and A79) Notch3 pathway was already a reality [75]. In a study from Machuca-Parra and Bigger-Allen et al., the authors use an anti-human Notch3 agonist antibody (A13, Genentech) that could be useful for patients with hypomorphic CADASIL mutations in Notch3 and also for others with small vessel disease conditions mechanistically linked to Notch3 loss of function [76]. Lateral meningocele syndrome (LMS) is a genetic disorder associated with *NOTCH3* mutations, and Yu et al. successfully reversed the

skeletal phenotype of LMS in male mice using an anti-Notch3 antibody (Genentech) [77]. A recent study used an anti-Notch3 antibody (Ab 28042, AVEO Pharmaceuticals), able to bind to both human and murine Notch3, as a drug treatment to reverse pulmonary hypertension (PH) in mouse hypoxia and rat Sugen-hypoxia models of PH [78]. Interestingly, a novel anti-Notch3 antibody-drug conjugate able to deliver an auristatin-based cytotoxic payload (PF-06650808) reached a phase I study for patients with breast cancer and other advanced solid tumors (NCT02129205). Unfortunately, the study was prematurely terminated as a business decision prior to the start of the dose expansion stage.

Despite promising preclinical results, targeting Notch3 signaling has not reached the clinic yet, but with the advent of new evidence involving Notch3 and disease, the development and use of cell-specific Notch3 inhibitors could be a useful strategy to tackle fibrosis.

5. Discussion and Conclusions

Fibrosis is characterized by excessive ECM deposition, which leads to scarring and organ failure. Fibrosis can affect many organs, and 45% of all deaths worldwide are estimated to result from a fibrotic disease. Unfortunately, there are no treatments capable of curing any of them despite the big efforts to understand their physiopathology. Numerous cellular and molecular mechanisms mediate fibrogenesis, such as the Notch pathway, a highly conserved cell-cell communication signaling pathway between neighbouring cells in which physical contact is needed. The role of Notch3 in some diseases is clear, like in CADASIL, where Notch3 mutations are strongly associated with its development. Moreover, the Notch3 receptor has other unique features among the Notch receptors (as shown in 3. Implication of Notch3 in the development of organ fibrosis), and its involvement in fibrogenesis is doubtless.

The use of global Notch3-knockout (KO) mice has allowed us to demonstrate the involvement of Notch3 signaling in the development of fibrosis in the lung and kidney in vivo. However, this tool has a significant caveat: Notch3 signaling is deleted systemically in every cell type and at every stage of development. Since the Notch pathway regulates many different biological processes depending on the cell type and the context, it is difficult to pinpoint the precise role of Notch3 in fibrogenesis using the Notch3 KO mouse line. To solve this problem, conditional Notch3 knockout mice under specific drivers are prime for targeting concrete cell types. In our laboratory, we have already generated various transgenic mouse lines in which we are able to delete Notch3 in a cell-specific manner, giving us the opportunity to accurately explore the role of Notch3 signaling in fibroblast activation.

It would also be interesting to assess Notch3 levels across diseases and cell types on the lookout for new therapeutic windows targeting Notch3 signaling in each fibrotic disease. A relation between upregulated Notch3 mRNA and protein levels with moderate and severe fibrosis stages has been shown in the liver. The fact that no significant relation is observed between mild patients and patients without liver fibrosis shows how the Notch3 pathway could act differently in different stages of fibrotic diseases. However, this is the only example regarding Notch3 levels and fibrotic disease stage and further investigation is needed to establish a clear association.

Despite previous evidence indicating a role for Notch3 in fibrotic disease, targeting Notch3 signaling is an arduous task due to the inherent complexity of the Notch pathway: (1) There are numerous Notch target genes, including the Hes and Hey family, and there is no clear pattern to which gene is activated by which NICD, (2) Each receptor may be activated by any Notch ligand and, depending on the ligand, this activation could lead to different molecular outcomes, (3) The Notch pathway can interact with other molecular pathways [79], (4) Instead of activating them (trans/cisactivation), ligands can also inhibit receptor activation (trans/cis-inhibition) [80]. All of this partially accounts for the cell and context dependency of Notch signaling and the paradoxical role of Notch3 in fibrotic disease, where both upregulation (liver, kidney, skin, pancreas, and lung) and downregulation (heart) of the pathway can trigger the development of fibrosis.

In general, studying the potential interactions between the Notch3 pathway and other signaling pathways and delving deeply into the mechanism involved in Notch3 activation in each context could shed light on new molecular mechanisms involved in fibrosis development and progression. This could help identify specific target genes associated with the development or progression of a fibrotic disease, which could be valuable in the design of new antifibrotic drugs.

Depending on the cellular and molecular context, Notch3 could regulate proliferation, migration and/or myofibroblast differentiation and play a role in inflammation and tissue regeneration. Therefore, since each cell type could have different roles in the context of injury, targeting Notch3 signaling in a cell-specific fashion is also highly important in the clinical setting. Of note, our previous publication, showed that the decrease in lung fibrosis in Notch3 deficient lungs is not due to a reduction in inflammation, supporting its role in myofibroblast activation [75]. This also

matches the observations made in the kidney, where Notch3 in the inflammatory component was not crucial for the development of fibrosis [46].

As described above, after binding to a ligand, the NICD is cleaved by a γ-secretase. Many of the clinical trials are focused specifically on the use of γ-secretase inhibitors. Therefore, they are not only inhibiting the signaling of all Notch receptors but also other pathways in which γ-secretases are involved. This and the fact that these treatments are not done in a cell-specific manner could explain why pan-Notch inhibitors often fail, leading to adverse effects.

In general, Notch3 plays an important role in tissue fibrogenesis at different levels and in different organs (Figure 2). Upon tissue injury, there is an immune response followed by the activation of fibroblasts, leading to myofibroblast differentiation and ECM production with the aim of healing the wound and regenerating the tissue. However, dysregulation in this process could lead to chronic fibrosis. The Notch3 could play a pivotal role not only in the process of inflammation but also in myofibroblast differentiation and ECM production in different organs. However, no clear results have been shown in every tissue.

In conclusion, there is still much to do to fully elucidate the implication of Notch3 signaling as a general mechanism in developing fibrosis in different organs. The use of specific mouse transgenic lines targeting Notch3 in specific cell types is extremely important in this matter. Moreover, using anti-Notch3 tools in ex vivo platforms such as precissioncut tissue slices or in vitro using human fibroblasts and organoids will provide decisive results about its potential therapeutic value in human fibrosis. Indeed, we are still far from having an approved anti-Notch3 drug to treat fibrosis, let alone encased in a molecular structure that gives it the capacity to act on the desired cell type in a specific organ. However, this could be the best scenario for the treatment of fibrotic diseases. Therefore, more focus should be placed on the collaboration among disciplines, such as those studying different biological aspects of a disease and those developing new strategies and technologies that could be used to deliver specific drugs targeting specific cells in the desired organ.

Figure 2. Notch3 contribution to inflammation and fibrosis. The cartoon reflects the different stages from tissue injury to its resolution or its chronicicity. First, an insult to the tissue leads to immune cell recruitment to the site of injury and cytokine release (inflammation). This elicits fibroblast activation and myofibroblast differentiation, which leads to ECM production to give the tissue the scaffold for proper regeneration. However, a dysregulation in any of these steps leads to an excessive scar formation, resulting in the development of chronic fibrotic disease. Despite the scarcity of studies on skin fibrosis, Notch3 upregulation could impact inflammation in the liver and kidneys. Moreover, Notch3 upregulation also promotes tissue fibrosis in the liver, pancreas,

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and lung, while it seems to have the opposite effect in the heart. All in all, targeting Notch3 signaling could potentially reverse tissue fibrosis and thus give a new opportunity for organ regeneration.

Acknowledgments

We acknowledge the support of every member of our laboratory for support and feedback on the manuscript.

Author Contributions

Conceptualization, A.E.-Z., Z.B.-I. and A.P.-S.; writing—original draft preparation, A.E.-Z.; writing—review and editing, A.E.-Z., Z.B.-I., B.S. and A.P.-S.; supervision, A.P.-S. and B.S. All authors have read and agreed to the published version of the manuscript.

Ethics Statement

Not applicable.

Informed Consent Statement

Not applicable.

Funding

This work received no external funding. It was supported by the laboratory of A.P.-S. that is funded by the Institute for Lung Health, Justus Liebig University Giessen (Germany).

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

References

- 1. Pakshir P, Hinz B. The big five in fibrosis: Macrophages, myofibroblasts, matrix, mechanics, and miscommunication. Matrix Biol. 2018, 68, 81–93.
- 2. Weiskirchen R, Weiskirchen S, Tacke F. Organ and tissue fibrosis: Molecular signals, cellular mechanisms and translational implications. Mol. Aspects Med. 2019, 65, 2–15.
- 3. Henderson NC, Rieder F, Wynn TA. Fibrosis: From mechanisms to medicines. Nature 2020, 587, 555–566.
- 4. Murtha LA, Schuliga MJ, Mabotuwana NS, Hardy SA, Waters DW, Burgess JK, et al. The processes and mechanisms of cardiac and pulmonary fibrosis. Front. Physiol. 2017, 8, 777.
- 5. Qin L, Liu N, Bao CL, Yang DZ, Ma GX, Yi WH, et al. Mesenchymal stem cells in fibrotic diseases—The two sides of the same coin. Acta Pharmacol. Sin. 2023, 44, 268–287.
- 6. Biasin V, Crnkovic S, Sahu-Osen A, Birnhuber A, El Agha E, Sinn K, et al. PDGFRα and αSMA mark two distinct mesenchymal cell populations involved in parenchymal and vascular remodeling in pulmonary fibrosis. Am. J. Physiol. Lung Cell. Mol. Physiol. 2020, 318, L684–L697.
- 7. Distler JHW, Györfi AH, Ramanujam M, Whitfield ML, Königshoff M, Lafyatis R. Shared and distinct mechanisms of fibrosis. Nat. Rev. Rheumatol. 2019, 15, 705–730.
- 8. Kiyokawa H, Morimoto M. Notch signaling in the mammalian respiratory system, specifically the trachea and lungs, in development, homeostasis, regeneration, and disease. Dev. Growth Differ. 2020, 62, 67–79.
- 9. Zhou B, Lin W, Long Y, Yang Y, Zhang H, Wu K, et al. Notch signaling pathway: Architecture, disease, and therapeutics. Signal Transduct. Target Ther. 2022, 7, 95.
- 10. Metz CW, Bridges CB. Incompatibility of mutant races in Drosophila. Proc. Natl. Acad. Sci. USA 1917, 3, 673–678.
- 11. Blokzijl A, Dahlqvist C, Reissmann E, Falk A, Moliner A, Lendahl U, et al. Cross-talk between the Notch and TGF-β signaling pathways mediated by interaction of the Notch intracellular domain with Smad3. J. Cell Biol. 2003, 163, 723–728.
- 12. Totaro A, Castellan M, Di Biagio D, Piccolo S. Crosstalk between YAP/TAZ and Notch Signaling. Trends Cell Biol. 2018, 28, 560–573.
- 13. Wu J, Li W, Guo L, Zhao L, Sun S, Li H. The crosstalk between the Notch, Wnt, and SHH signaling pathways in regulating the proliferation and regeneration of sensory progenitor cells in the mouse cochlea. Cell Tissue Res. 2021, 386, 281–296.
- 14. Kopan R, Ilagan MaXG. The Canonical Notch Signaling Pathway: Unfolding the Activation Mechanism. Cell 2009, 137, 216–233.
- 15. Sanchez-Niño MD, Ortiz A. Notch3 and kidney injury: Never two without three. J. Pathol. 2012, 228, 266–273.
- 16. Xu X, Choi SH, Hu T, Tiyanont K, Habets R, Groot AJ, et al. Insights into autoregulation of Notch3 from structural and functional studies of its negative regulatory region. Structure 2015, 23, 1227–1235.
- 17. Choy L, Hagenbeek TJ, Solon M, French D, Finkle D, Shelton A, et al. Constitutive NOTCH3 signaling promotes the growth of basal breast cancers. Cancer Res. 2017, 77, 1439–1452.
- 18. Wang MM. CADASIL. In Handbook of Clinical Neurology; Elsevier: Amsterdam, The Netherlands, 2018; pp. 733–743.
- 19. Xiu M, Wang Y, Li B, Wang X, Xiao F, Chen S, et al. The role of Notch3 signaling in cancer stemness and chemoresistance: Molecular mechanisms and targeting strategies. Front. Mol. Biosci. 2021, 8, 694141.
- 20. Marcellin P, Kutala BK. Liver diseases: A major, neglected global public health problem requiring urgent actions and largescale screening. *Liver Int.* **2018**, 38, 2–6.
- 21. Valizadeh A, Sayadmanesh A, Asemi Z, Alemi F, Mahmoodpoor A, Yousefi B. Regulatory roles of the Notch signaling pathway in liver repair and regeneration: A novel therapeutic target. Curr. Med. Chem. 2021, 28, 8608–8626.
- 22. Xie G, Wang X, Wang L, Wang L, Atkinson RD, Kanel GC, et al. Role of differentiation of liver sinusoidal endothelial cells in progression and regression of hepatic fibrosis in rats. Gastroenterology 2012, 142, 918–927.
- 23. Tsuchida T, Friedman SL. Mechanisms of hepatic stellate cell activation. Nat. Rev. Gastroenterol. Hepatol. 2017, 14, 397– 411.
- 24. Wang SS, Tang XT, Lin M, Yuan J, Peng YJ, Yin X, et al. Perivenous stellate cells are the main source of myofibroblasts and cancer-associated fibroblasts formed after chronic liver injuries. Hepatology 2021, 74, 1578–1594.
- 25. Wynn T, Barron L. Macrophages: Master regulators of inflammation and fibrosis. Semin. Liver Dis. 2010, 30, 245–257.
- 26. Heymann F, Trautwein C, Tacke F. Monocytes and macrophages as cellular targets in liver fibrosis. Inflamm. Allergy Drug Targets 2009, 8, 307–318.
- 27. Sica A, Invernizzi P, Mantovani A. Macrophage plasticity and polarization in liver homeostasis and pathology. *Hepatology* 2014, 59, 2034–2042.
- 28. Bansal R, van Baarlen J, Storm G, Prakash J. The interplay of the Notch signaling in hepatic stellate cells and macrophages determines the fate of liver fibrogenesis. Sci. Rep. 2015, 5, 18272.
- 29. Chen YX. Notch3 regulates the activation of hepatic stellate cells. World J. Gastroenterol. 2012, 18, 1397.
- 30. Nijjar S. Notch receptor expression in adult human liver: A possible role in bile duct formation and hepatic neovascularization. Hepatology 2001, 34, 1184–1192.
- 31. Chen Y, Zheng S, Qi D, Zheng S, Guo J, Zhang S, et al. Inhibition of Notch signaling by a γ-secretase inhibitor attenuates hepatic fibrosis in rats. PLoS ONE 2012, 7, e46512.
- 32. Zhu C, Kim K, Wang X, Bartolome A, Salomao M, Dongiovanni P, et al. Hepatocyte Notch activation induces liver fibrosis in nonalcoholic steatohepatitis. Sci. Transl. Med. 2018, 10, eaat0344.
- 33. Ramachandran P, Dobie R, Wilson-Kanamori JR, Dora EF, Henderson BEP, Luu NT, et al. Resolving the fibrotic niche of human liver cirrhosis at single-cell level. *Nature* 2019, 575, 512–518.
- 34. Cong S, Liu Y, Li Y, Chen Y, Chen R, Zhang B, et al. MiR-571 affects the development and progression of liver fibrosis by regulating the Notch3 pathway. Sci. Rep. 2021, 11, 21854.
- 35. Moeller MJ, Kramann R, Lammers T, Hoppe B, Latz E, Ludwig-Portugall I, et al. New aspects of kidney fibrosis–from mechanisms of injury to modulation of disease. Front. Med. 2022, 8, 814497.
- 36. Edeling M, Ragi G, Huang S, Pavenstädt H, Susztak K. Developmental signaling pathways in renal fibrosis: The roles of Notch, Wnt and Hedgehog. Nat. Rev. Nephrol. 2016, 12, 426–439.
- 37. Djudjaj S, Chatziantoniou C, Raffetseder U, Guerrot D, Dussaule JC, Boor P, et al. Notch-3 receptor activation drives inflammation and fibrosis following tubulointerstitial kidney injury. J. Pathol. 2012, 228, 286–299.
- 38. Xiao Z, Zhang J, Peng X, Dong Y, Jia L, Li H, et al. The Notch γ-secretase inhibitor ameliorates kidney fibrosis via inhibition of TGF-β/Smad2/3 signaling pathway activation. Int. J. Biochem. Cell Biol. 2014, 55, 65–71.
- 39. Han SH, Wu MY, Nam BY, Park JT, Yoo TH, Kang SW, et al. PGC-1α protects from Notch-induced kidney fibrosis development. J. Am. Soc. Nephrol. 2017, 28, 3312–3322.
- 40. Xue C, Mei CL. Polycystic Kidney Disease and Renal Fibrosis. In Advances in Experimental Medicine and Biology; Springer: Singapore, 2019; Volume 1165, pp. 81–100.
- 41. Djudjaj S, Kavvadas P, Prakoura N, Bülow RD, Migeon T, Placier S, et al. Activation of Notch3 in renal tubular cells leads to progressive cystic kidney disease. Int. J. Mol. Sci. 2022, 23, 884.
- 42. Guerrot D, François A, Boffa JJ, Boulos N, Hanoy M, Legallicier B, et al. Nephroangiosclerosis in cerebral autosomal dominant arteriopathy with subcortical infarcts and leukoencephalopathy: Is NOTCH3 mutation the common culprit? Am. J. Kidney Dis. 2008, 52, 340–345.
- 43. Boulos N, Helle F, Dussaule JC, Placier S, Milliez P, Djudjaj S, et al. Notch3 is essential for regulation of the renal vascular tone. Hypertension 2011, 57, 1176–1182.
- 44. Huang S, Park J, Qiu C, Chung KW, Li SY, Sirin Y, et al. Jagged1/Notch2 controls kidney fibrosis via Tfam-mediated metabolic reprogramming. PLoS Biol. 2018, 16, e2005233.
- 45. Brandt S, Ballhause TM, Bernhardt A, Becker A, Salaru D, Le-Deffge HM, et al. Fibrosis and immune cell infiltration are separate events regulated by cell-specific receptor Notch3 expression. J. Am. Soc. Nephrol. 2020, 31, 2589–2608.
- 46. Do NN, Eming SA. Skin fibrosis: Models and mechanisms. Curr. Res. Transl. Med. 2016, 64, 185–193.
- 47. He T, Bai X, Jing J, Liu Y, Wang H, Zhang W, et al. Notch signal deficiency alleviates hypertrophic scar formation after wound healing through the inhibition of inflammation. Arch. Biochem. Biophys. 2020, 682, 108286.
- 48. Distler A, Lang V, Del Vecchio T, Huang J, Zhang Y, Beyer C, et al. Combined inhibition of morphogen pathways demonstrates additive antifibrotic effects and improved tolerability. Ann. Rheum. Dis. 2014, 73, 1264–1268.
- 49. Tao Y, Yang Q, Wang L, Zhang J, Zhu X, Sun Q, et al. β-Catenin activation in hair follicle dermal stem cells induces ectopic hair outgrowth and skin fibrosis. J. Mol. Cell. Biol. 2019, 11, 26-38.
- 50. Zmorzyn'ski S, Wojcierowska-Litwin M, Kowal M, Michalska-Jakubus M, Styk W, Filip AA, et al. NOTCH3 T6746C and TP53 P72R Polymorphisms Are Associated with the Susceptibility to Diffuse Cutaneous Systemic Sclerosis. Biomed. Res. Int. 2020, 1, 1–9.
- 51. Li L, Zuo X, Liu D, Luo H, Zhu H. The profiles of miRNAs and lncRNAs in peripheral blood neutrophils exosomes of diffuse cutaneous systemic sclerosis. J. Dermatol. Sci. 2020, 98, 88–97.
- 52. Condorelli AG, Nobili R, Muglia A, Scarpelli G, Marzuolo E, De Stefanis C, et al. Gamma-Secretase Inhibitors Downregulate the Profibrotic NOTCH Signaling Pathway in Recessive Dystrophic Epidermolysis Bullosa. J. Investig. Dermatol. 2024, 144, 1522–1533.e10.
- 53. Negri VA, Logtenberg ME, Renz LM, Oules B, Walko G, Watt FM. Delta-like 1-mediated cis-inhibition of Jagged1/2 signaling inhibits differentiation of human epidermal cells in culture. Sci. Rep. 2019, 9, 10825.
- 54. Roth GA, Mensah GA, Johnson CO, Addolorato G, Ammirati E, Baddour LM, et al. Global burden of cardiovascular diseases and risk factors, 1990–2019: Update from the GBD 2019 study. J. Am. Coll. Cardiol. 2020, 76, 2982–3021.
- 55. Mewton N, Liu CY, Croisille P, Bluemke D, Lima JA. Assessment of myocardial fibrosis with cardiovascular magnetic resonance. J. Am. Coll. Cardiol. 2011, 57, 891–903.
- 56. Li L, Zeng H, Chen JX. Apelin-13 increases myocardial progenitor cells and improves repair postmyocardial infarction. Am. J. Physiol. Heart Circ. Physiol. 2012, 303, H605–H618.
- 57. Li L, Zeng H, Hou X, He X, Chen JX. Myocardial injection of apelin-overexpressing bone marrow cells improves cardiac repair via upregulation of Sirt3 after myocardial infarction. PLoS ONE 2013, 8, e71041.
- 58. Del Gaudio F, Liu D, Andaloussi Mäe M, Braune EB, Hansson EM, Wang QD, et al. Left ventricular hypertrophy and metabolic resetting in the Notch3-deficient adult mouse heart. Sci. Rep. 2023, 13, 15022.
- 59. Chen JX, Chen ST, Tao YK. Cardiac pericyte is promising target for ischemic heart diseases: Role of Notch3. Int. J. Cardiol. 2017, 246, 57.
- 60. Shi J, Xiao P, Liu X, Chen Y, Xu Y, Fan J, et al. Notch3 modulates cardiac fibroblast proliferation, apoptosis, and fibroblast to myofibroblast transition via negative regulation of the RhoA/ROCK/Hif1α axis. Front. Physiol. 2020, 11, 669.
- 61. Zhang M, Pan X, Zou Q, Xia Y, Chen J, Hao Q, et al. Notch3 ameliorates cardiac fibrosis after myocardial infarction by inhibiting the TGF-β1/Smad3 pathway. Cardiovasc. Toxicol. 2016, 16, 316–324.
- 62. Huang C, Iovanna J, Santofimia-Castaño P. Targeting Fibrosis: The Bridge That Connects Pancreatitis and Pancreatic Cancer. Int. J. Mol. Sci. 2021, 22, 4970.
- 63. Bhanot U, Köhntop R, Hasel C, Möller P. Evidence of Notch pathway activation in the ectatic ducts of chronic pancreatitis. J. Pathol. 2008, 214, 312–319.
- 64. Wang D, Xin L, Lin JH, Liao Z, Ji JT, Du TT, et al. Identifying miRNA-mRNA regulation network of chronic pancreatitis based on the significant functional expression. Medicine 2017, 96, e6668.
- 65. Mizrahi JD, Surana R, Valle JW, Shroff RT. Pancreatic cancer. Lancet 2020, 395, 2008–2020.
- 66. Mahadevan D, Von Hoff DD. Tumor-stroma interactions in pancreatic ductal adenocarcinoma. Mol. Cancer. Ther. 2007, 6, 1186–1197.
- 67. Song H, Zhang Y. Regulation of pancreatic stellate cell activation by Notch3. BMC Cancer 2018, 18, 36.
- 68. Eto K, Kawakami H, Kuwatani M, Kudo T, Abe Y, Kawahata S, et al. Human equilibrative nucleoside transporter 1 and Notch3 can predict gemcitabine effects in patients with unresectable pancreatic cancer. Br. J. Cancer 2013, 108, 1488–1494.
- 69. Xu K, Nieuwenhuis E, Cohen BL, Wang W, Canty AJ, Danska JS, et al. Lunatic Fringe-mediated Notch signaling is required for lung alveogenesis. Am. J. Physiol. Lung Cell Mol. Physiol. 2010, 298, L45–L56.
- 70. Hu B, Wu Z, Bai D, Liu T, Ullenbruch MR, Phan SH. Mesenchymal deficiency of Notch1 attenuates bleomycin-induced pulmonary fibrosis. Am. J. Pathol. 2015, 185, 3066–3075.
- 71. Xie T, Wang Y, Deng N, Huang G, Taghavifar F, Geng Y, et al. Single-cell deconvolution of fibroblast heterogeneity in mouse pulmonary fibrosis. Cell Rep. 2018, 22, 3625–3640.
- 72. Reyfman PA, Walter JM, Joshi N, Anekalla KR, McQuattie-Pimentel AC, Chiu S, et al. Single-cell transcriptomic analysis of human lung provides insights into the pathobiology of pulmonary fibrosis. Am. J. Respir. Crit. Care Med. 2019, 199, 1517– 1536.
- 73. Vera L, Garcia-Olloqui P, Petri E, Viñado AC, Valera PS, Blasco-Iturri Z, et al. Notch3 deficiency attenuates pulmonary fibrosis and impedes lung-function decline. Am. J. Respir. Cell Mol. Biol. 2021, 64, 465–476.
- 74. Lai JM, Zhang X, Liu FF, Yang R, Li SY, Zhu LB, et al. Redox-sensitive MAPK and Notch3 regulate fibroblast differentiation and activation: A dual role of ERK1/2. Oncotarget 2016, 7, 43731.
- 75. Li K, Li Y, Wu W, Gordon WR, Chang DW, Lu M, et al. Modulation of Notch signaling by antibodies specific for the extracellular negative regulatory region of NOTCH3. J. Biol. Chem. 2008, 283, 8046–8054.
- 76. Machuca-Parra AI, Bigger-Allen AA, Sanchez AV, Boutabla A, Cardona-Vélez J, Amarnani D, et al. Therapeutic antibody targeting of Notch3 signaling prevents mural cell loss in CADASIL. J. Exp. Med. 2017, 214, 2271–2282.
- 77. Yu J, Siebel CW, Schilling L, Canalis E. An antibody to Notch3 reverses the skeletal phenotype of lateral meningocele syndrome in male mice. J. Cell Physiol. 2020, 235, 210–220.
- 78. Zhang Y, Hernandez M, Gower J, Winicki N, Morataya X, Alvarez S, et al. Thistlethwaite PA. JAGGED-NOTCH3 signaling in vascular remodeling in pulmonary arterial hypertension. Sci. Transl. Med. 2022, 14, eabl5471.
- 79. Sachan N, Sharma V, Mutsuddi M, Mukherjee A. Notch signaling: Multifaceted role in development and disease. FEBS J. 2024, 291, 3030–3059.
- 80. Nandagopal N, Santat LA, Elowitz MB. Cis-activation in the Notch signaling pathway. Elife 2019, 8, e37880.